



## **Test Report**

Efficacy of A New JM Nanocomposite Material in Inhibiting Enterovirus  
Suspension Cellular Infection

### **Test Reagent**

New JM nanocomposite material

### **Project Commissioner**

JM Material Technology Inc.

### **Project Implementation Unit**

Cell Biology Laboratory, Cathay Medical Research Institute, Department  
of Medical Research, Cathay General Hospital

### **Testing Laboratory**

Virology Laboratory, Cathay General Hospital, Sijhih Branch

### **Project Personnel**

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Qing-Dong Ling

Signature: \_\_\_\_\_



## Abstract

**Title:** Efficacy of A New JM Nanocomposite Material in Inhibiting Enterovirus Suspension Cellular Infection

**Experiment design:** This study tested the efficacy of a new JM nanocomposite material in inhibiting enterovirus suspension cellular infection. A TCID<sub>50</sub> assay was used in an antiviral test to observe the cytopathic effect of infected cells in JM nanomaterials treated with a virus-enriched culture fluid to calculate the efficacy of JM nanomaterials inhibiting virus.

**Test reagent:** New JM nanocomposite material

**Reagent vendor:** JM Material Technology Inc., 5F.-3, No. 40-2, Sec. 1, Minsheng N Rd., Guishan Township, Taoyuan County



## Test Content

### Experiment Materials

Virus strain source:

Enterovirus echovirus type 11 sourced from a College of American Pathologists proficiency-testing specimen.

Host cell:

LLC-MK2 cells (BCRC 60092) procured from the Bioresource Collection and Research Center, Taiwan, R.O.C.

### Experimental Methods:

#### 1. Cell culture

- (a) Inoculate the cell strains in a 24-well culture plate.
- (b) Incubate cells minimum essential medium (MEM) supplemented with 8% fetal bovine serum at 36 °C with 5% CO<sub>2</sub> for 48 h until fully grown.
- (c) Discard the culture fluid, rinse twice with phosphate buffer saline (PBS), and set aside.

#### 2. Virus preparation

- (a) Inoculate the virus in culture tubes containing cell strains.
- (b) Incubate cells in MEM at 36 °C with 5% CO<sub>2</sub> for 48 h until cytopathy occurs.
- (c) Scrape off the cells and precipitate cells by centrifugation at 6000 rpm for 2 min.
- (d) The supernatant is collected as the virus suspension.
- (e) Add 1080 uL of MEM to 120 uL of the virus suspension and dilute it at a ratio of 1:10.
- (f) Add 120 uL of the above dilution to 1080 uL of MEM and perform a 10-fold serial dilution.

Number	1	2	...	6	7	8	9
MEM (+ Trypsin)	900	900	...	900	900	900	900
Virus suspension	100	0	...	0	0	0	0
Serial dilution							
Final volume	900	900	...	900	900	900	1000
Final concentration	10 <sup>-1</sup>	10 <sup>-2</sup>	...	10 <sup>-6</sup>	10 <sup>-7</sup>	10 <sup>-8</sup>	10 <sup>-9</sup>



### 3. TCID<sub>50</sub> Assay

#### Control group

Use a 24-well plate with seeded cells. Leave the 4 culture tubes in Column 1 untreated as the control, and treat Column 2 to 6 with the enterovirus by adding 200 uL of virus suspension at 10<sup>5</sup>-, 10<sup>6</sup>-, 10<sup>7</sup>-, 10<sup>8</sup>-, and 10<sup>9</sup>-fold dilutions, respectively.

#### Experimental group

- Prepare a 5-fold dilution by adding 100 uL of the virus suspension to 400 uL of MEM.
- Prepare a 10-fold serial dilution by adding 50 uL of the dilution to 450 uL of MEM.
- Prepare 1.25% disinfectant (75 uL of disinfectant + 5925 uL of MEM) and add 450 uL of the disinfectant to each of the above dilution.
- Prepare 450 uL of the 1.25% disinfectant, adding it to 450 uL of virus-free MEM for the JM toxicity test.

Number	BC	B1	B2	B3	B4	B5
MEM (+ Trypsin)	450	450	450	450	450	450
Virus suspension	0	100	0	0	0	0
Serial dilution		50	50	50	50	50
1.25% Disinfectant	450	450	450	450	450	450
Final Volume	900	900	900	900	900	900
Final concentration of virus	0	10 <sup>-1</sup>	10 <sup>-2</sup>	10 <sup>-3</sup>	10 <sup>-4</sup>	10 <sup>-5</sup>
Final concentration of disinfectant	0.625%	0.625%	0.625%	0.625%	0.625%	0.625%

Discarding

- Expose the dilution to UV for 1 h at room temperature.
- Among the cell strains in the 24-well culture plate, inoculate the four wells in Column 1 for the JM toxicity test; inoculate the remaining five columns with 200 uL of 10<sup>1</sup>-, 10<sup>2</sup>-, 10<sup>3</sup>-, 10<sup>4</sup>-, and 10<sup>5</sup>-fold diluted virus suspension that is treated with 0.625% of the JM nanomaterial.



Allow both the experimental group and control group to be infected for 1 h at 36 °C and 5% CO<sub>2</sub>, and shake them every 20 min. Add MEM (+Trypsin) to each culture tube, incubate at 36 °C with 5% CO<sub>2</sub>, and observe daily for the number of tubes displaying cell pathology. Add 1 mL of 4% formaldehyde and leave them to stand at room temperature for 1 h. Rinse them twice with tap water, add 1 mL of 0.5% crystal violet, and leave them to stand at room temperature for 5 min.

#### 4. Interpretation and Calculation

- (a) The Reed–Muench method was used to calculate TCID<sub>50</sub>.
- (b) Formula for calculating viral inhibitory efficacy:  
percentage of inhibition =  $[1 - 10^{-(\text{viral load of the control group} - \text{viral load of the experimental group})}] \times 100$



## Test Results

### Enterovirus

Group	Viral load (Log <sub>10</sub> TCID <sub>50</sub> )		
	1st	2nd	3rd
Virus strains	6.7	7.5	6.7
Virus strains+JM	4.5	4.3	4.7
Cell strains	None	None	None

Calculation of viral inhibitory efficacy:

Substituting the mean of the three test results obtained the following results:

Enterovirus inhibition percentage =  $[1 - 10^{-(7.0-4.5)}] \times 100 = 99.68$

## Conclusion

The experiment results show that a 0.625% concentration of the JM nanomaterials inhibit cellular infection of enterovirus. The percentage of viral inhibition was **99.68%**.